

Stata commands to use

Before following the tips in this document, please import your excel document into Stata. Below examples are based off Research Office Biostatistician data.

Tips

Syntax is the grammar of Stata commands. Stata is a command-based language. Most Stata commands are verbs. They tell Stata to do something: summarize, tabulate, regress, etc.

Basic Stata command syntax:

The name of the command followed by a list of variables and instructions. For the specific syntax for a particular command click help. For example, to do a regression, regress is the name of the command. The first variable listed (`logTimeClose`) is the outcome variable the others (`CHRONICINFECTION` `Sex` `BCereus` `BMI`) are predictor variables. The `i.` and `##` are specific instructions on how to work with the variables. After a comma, Stata allows you to use "options" which modify how the command is used. `vce(robust)` tells Stata to calculate the robust standard error. Help will list all the available options for a command.

```
. regress logTimeClose i.CHRONICINFECTION##i.Sex i.BCereus BMI, vce(robust)
```

Some commands can be combined using brackets. For example, a scatter graph and a line fit graph are combined on a single two-way graph.

```
. twoway (scatter logTimeClose Age, sort) (lfit logTimeClose Age, sort)
```

You can access the Help Tab from the command line. For example, to identify the collinearity diagnostics command `collin`...

```
. help collin
```

If you do not know the specific Stata command, use:

```
. search collinearity
```

In Stata, you can keep a log file of all commands and results (Graphical output is exempt). You can also copy and paste these to a work file. If you want to provide annotation, put a `*` before your comment as shown below:

```
. * Here's how to use help from the command line
```

```
. help collin
```

If the comment is long, use `/*...*/`. Example below:

```
/* Here I will recode BMI into categories using the  
ABS categories. Personally, I don't think we should use categories */
```

Using `/* ... */` comments can be appended directly to commands. The `recode` command is used to create categories. `9/18` means values 9 to 18, `1` is the numerical value of the category, and `Underweight` is the label for the category. If the label has a space in it you have to put it in quotations. Once the categories are defined the option is to generate a new variable called `BMI_Cat`.

```
recode BMI (9/18 = 1 Underweight) (19/24 = 2 Healthy) (25/29 = 3 Overweight) (30/39 = 4  
Obese) (40/max = 5 "Extremely Obese"), gen(BMI_Cat) /* ABS categories */
```

A command can be conditional by using `if` along with other logical operators. For example, if you wanted to do a regression of `BMI` as the outcome and `education` as the predictor but only for single parent households with more than one child.

```
regress BMI education if parents==1 & nchildren > 1
```

Stata's Logical and Relational Operators

```
& and
```

```
| or
```



```
! not
~ not
> greater than
< less than
>= greater or equal
<= less than or equal
== equal
!= not equal
```

If you want to repeat a command for each subgroup of a variable use `by`. It usually comes before the command separated by a colon: For example, to create a table of statistics for `TIMETOCLOSURE` for each category of `BCereus` (presence or absence of infection with this bacteria).

```
. by BCereus, sort: tabstat TIMETOCLOSURE, stat(mean sd p25 p50 p75)
```

Note: `by` usually only works if `BCereus` is sorted, hence using the option `sort`. You can do both together with `bysort`.

```
. bysort BCereus: tabstat TIMETOCLOSURE, stat(mean sd p25 p50 p75)
```

`by` can also be used as an option with some commands using the syntax `by(varname)`, For example:

```
. tabstat TIMETOCLOSURE, by(BCereus) stat(mean sd p25 p50 p75)
```

This gives a neater output and does not require the subgrouping variable to be sorted.

In addition to the resources listed in the “Introduction to Using Stata 15 at Gold Coast Health”, <http://wlm.userweb.mwn.de/Stata/wstatbas.htm> is an informative website to follow.

Summary Statistics and Graphics

To start understanding your data, run some basic summary statistics and graphs to visualize the data.

```
. summarize TIMETOCLOSURE, detail
```

```
TIME TO CLOSURE
-----
Percentiles   Smallest
1%           19
5%           60.5      34
10%          72         52   Obs           71
25%          180        60.5   Sum of Wgt.    71

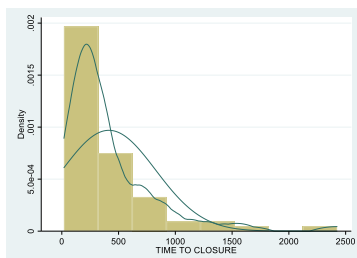
50%          288
Largest      Std. Dev.   411.4733
75%          560        1224
90%          843.7      1446   Variance       169310.2
95%          1224       1629   Skewness       2.437968
99%          2424       2424   Kurtosis       10.5143
```

```
. histogram TIMETOCLOSURE
(bin=8, start=19, width=300.625)
```



Add in the options normal and kdensity superimpose a normal curve and an empirical estimate of the frequency distribution.

```
. histogram TIMETOCLOSURE, normal kdensity
(bin=8, start=19, width=300.625)
```



The distribution above does not look normal (no left tail and skewed to the right). To do a formal test of normality

```
. swilk TIMETOCLOSURE
```

Shapiro-Wilk W test for normal data

Variable	Obs	W	V	z	Prob>z
TIMETOCLOS~E	71	0.74797	15.692	5.992	0.00000

As expected, the probability that the underlying data from which this sample was taken is normal and is tiny. The data is significantly different from being normally distributed.

Generating variables and cleaning and organising your data

[Data tab, Create or Change Data].

```
. gen AgeDec = Age/10
```

Once a variable is generated it can be modified with the replace command.

```
. replace AgeDec = 8 if AgeDec>8
```

The BMI variable, in this example was in red because it was text due to an “NA” appearing in the data. First, we remove the NA by replacing it with nothing.

```
. replace BMI = "" in 27
(1 real change made)
```

There are two ways to convert tie text variable to a numerical variable. First, generate a new variable that is numerical using the real command.

```
. gen BMIr =real(BMI)
(1 missing value generated)
```

Second, convert the original text variable to a numerical variable using destring.

```
. destring BMI, replace
BMI: all characters numeric; replaced as double
(1 missing value generated)
```

The other text variable is Sex. There are two ways to convert this to numeric.

First, use encode. [Data tab, Create or Change Data, Other Variable Transformation Commands].

```
. encode Sex, gen(Sex12)
```

New variables are added to the bottom of the variable list but can be moved to a more convenient location with the order command.

```
. order Sex12, after( Sex)
```

Second, replace the values and convert to numeric.

```
. replace Sex="0" if Sex=="F"
(43 real changes made)

. replace Sex="1" if Sex=="M"
(28 real changes made)

. destring Sex, replace
Sex: all characters numeric; replaced as byte
```

Values in numeric variables can be labelled.

```
. label define Gender 0 "F" 1 "M"
. label values (Sex) Gender
```

NOTE: Using encode automatically labelled the new variable Sex12 with F and M. However, it encoded the original Fs and Ms with 1s and 2s (not, as I had hoped, with 0s and **1s**).

Basic data analysis

Association between two categorical variables.

```
. tabulate CHRONICINFECTION Bonehealing, chi2 exact
```

Enumerating sample-space combinations:

```
stage 5: enumerations = 1
stage 4: enumerations = 2
stage 3: enumerations = 5
stage 2: enumerations = 16
stage 1: enumerations = 0
```

CHRONIC INFECTION	Bone healing					Total
	1	2	3	4	5	
0	38	2	5	3	1	49
1	14	3	2	3	0	22
Total	52	5	7	6	1	71

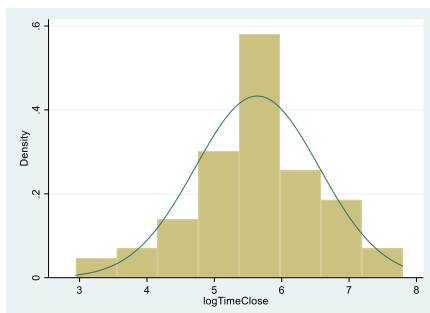
```
Pearson chi2(4) = 3.8521 Pr = 0.426
Fisher's exact = 0.402
```

```
. tabulate Bonehealing CHRONICINFECTION, chi2 exact
```

Log transformation:

```
. gen logTimeClose =ln( TIMETOCLOSURE)

. histogram logTimeClose, normal
(bin=8, start=2.9444389, width=.60609192)
```



```
. swilk logTimeClose
```

```
Shapiro-Wilk W test for normal data
```

Variable	Obs	W	V	z	Prob>z
logTimeClose	71	0.98790	0.753	-0.616	0.73112

Compare time to closure by Chronic infection. Non-parametric ranksum (Man-Whitney U).

```
. ranksum TIMETOCLOSURE, by( CHRONICINFECTION)
```

Two-sample Wilcoxon rank-sum (Mann-Whitney) test

CHRONICINF~N	obs	rank sum	expected
0	49	1708.5	1764
1	22	847.5	792
combined	71	2556	2556

unadjusted variance	6468.00
adjustment for ties	-3.69
adjusted variance	6464.31

Ho: TIMETO~E(CHRONI~N==0) = TIMETO~E(CHRONI~N==1)
z = -0.690
Prob > |z| = 0.4900

t-test of log transformed data:

```
. ttest logTimeClose, by( CHRONICINFECTION)
```

Two-sample t test with equal variances

Group	Obs	Mean	Std. Err.	Std. Dev.	[95% Conf. Interval]	
0	49	5.563024	.1269281	.8884966	5.307818	5.81823
1	22	5.802506	.2113022	.9910952	5.363079	6.241933
combined	71	5.63723	.1093155	.9211087	5.419207	5.855253
diff		-.2394812	.2363458		-.7109781	.2320157
diff = mean(0) - mean(1)				t =	-1.0133	
Ho: diff = 0				degrees of freedom =	69	

Ha: diff < 0 Ha: diff != 0 Ha: diff > 0
Pr(T < t) = 0.1572 Pr(|T| > |t|) = 0.3145 Pr(T > t) = 0.8428

The tabstat command can be used to compare summary statistics across categories of a categorical variable. For example below, time to closure by infection with B. Cereus.

```
. tabstat TIMETOCLOSURE, by( BCereus) stat(mean sd p25 p50 p75)
```

Summary for variables: TIMETOCLOSURE
by categories of: BCereus (B.Cereus)

BCereus	mean	sd	p25	p50	p75
0	280.7451	196.3073	144	221	384
1	757.05	592.3052	264	720	1068
Total	414.9155	411.4733	180	288	560

To estimate statistics like means or proportions, use:

[Statistics / Summaries, tables, and tests / Summary and descriptive statistics / Proportions]

```
. proportion Sex12
```

Proportion estimation Number of obs = 71

	Proportion	Std. Err.	Logit	[95% Conf. Interval]
--	------------	-----------	-------	----------------------

Sex12				
F		.6056338	.0579997	.4861713
M		.3943662	.0579997	.286321
				.713679
				.5138287

Then test against a specified value, example below.

Statistics / Summaries, tables, and tests / Classical tests of hypotheses / Proportion test

```
. prtest Sex12 == 0.5
Sex12 is not a 0/1 variable
r(450);
```

NOTE: There is an Error message because this test was expecting the variable to consist of 0s or 1s and as noted when Sex12 was encoded. Redoing it with the Sex variable will work.

```
. prtest Sex == 0.5
```

One-sample test of proportion		Number of obs	=	71
Variable		Mean	Std. Err.	[95% Conf. Interval]
Sex		.3943662	.0579997	.2806889 .5080435
p = proportion(Sex)				z = -1.7802
Ho: p = 0.5				
Ha: p < 0.5		Ha: p != 0.5		Ha: p > 0.5
Pr(Z < z) = 0.0375		Pr(Z > z) = 0.0750		Pr(Z > z) = 0.9625

To graph your data, use:

```
Graphics / Twoway graph (scatter, line, etc)
```

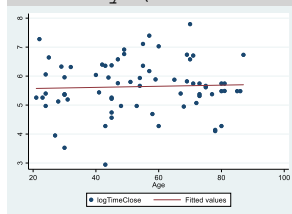
Two-way is the most used graphics command. Use the help tab to find out more about two-way.

It is important to include the sort option

```
. twoway (scatter logTimeClose Age, sort)
```

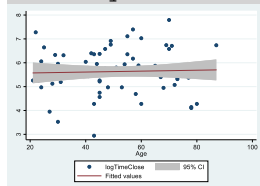
You can combine graphs on the same graph, For example, adding a linear best fit line (lfit).

```
. twoway (scatter logTimeClose Age, sort) (lfit logTimeClose Age, sort)
```



With confidence intervals

```
. twoway (scatter logTimeClose Age, sort) (lfitci logTimeClose Age, sort)
```

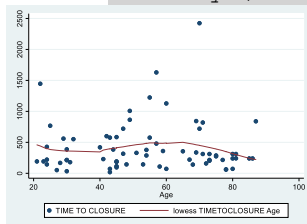


```
. twoway (scatter TIMETOCLOSURE Age, sort) (lfitci TIMETOCLOSURE Age, sort)
```



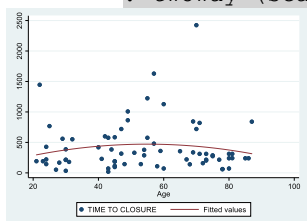
Lowess smothing

```
. twoway (scatter TIMETOCLOSURE Age, sort) (lowess TIMETOCLOSURE Age, sort)
```



Quadratic

```
. twoway (scatter TIMETOCLOSURE Age, sort) (qfit TIMETOCLOSURE Age, sort)
```



Linear regression:

```
Statistics / Linear models and related / linear regression
```

In terms of the syntax, write the command followed by the dependent (outcome) variable and then the list of independent (predictor) variables. Following a comma, add in options.

NOTE: the results below for CHRONICINFECTION are the same as for the t-test, P=0.314, so chronic infection does not significantly affect log time to closure.

NOTE: The i. before CHRONICINFECTION. This tells Stata to treat the variable as a set of indicator variables (a 0 or 1 for each category of a categorical variable, except the base category to which the other categories are compared). If you don't use the i. the variable is treated like a scale variable assuming equal distances between each category. If there are only two categories in the categorical variable there would only be one indicator variable so the analyses would be equivalent. It is best to routinely use the i. for any categorical variable.

```
. regress logTimeClose i.CHRONICINFECTION
```

Source	SS	df	MS	Number of obs	=	71
Model	.870769496	1	.870769496	F(1, 69)	=	1.03
Residual	58.5201186	69	.848117661	Prob > F	=	0.3145
				R-squared	=	0.0147
				Adj R-squared	=	0.0004
Total	59.3908881	70	.848441259	Root MSE	=	.92093

logTimeClose	Coef.	Std. Err.	t	P> t	[95% Conf. Interval]
1.CHRONICINFECTION	.2394812	.2363458	1.01	0.314	-.2320157 .7109781
_cons	5.563024	.1315619	42.28	0.000	5.300566 5.825483

Predictor variables are added below.

NOTE: Bone healing is presented, with coefficients for each category, these being relative to the base category which is category 1.

```
. regress logTimeClose i.CHRONICINFECTION Age BMI i.Sex i.Bonehealing i.BCereus
```

Source	SS	df	MS	Number of obs	=	70
Model	18.5791296	9	2.06434773	F(9, 60)	=	3.04
Residual	40.7421598	60	.679035996	Prob > F	=	0.0046
				R-squared	=	0.3132
				Adj R-squared	=	0.2102

```
Total | 59.3212893      69 .859728831  Root MSE      =      .82404
```

logTimeClose	Coef.	Std. Err.	t	P> t	[95% Conf. Interval]	
1.CHRONICINFECTION	.0823017	.2224124	0.37	0.713	-.3625892	.5271927
Age	-.0006995	.0055025	-0.13	0.899	-.0117062	.0103071
BMI	.0227162	.0172758	1.31	0.194	-.0118405	.057273
Sex						
M	-.1788183	.2194999	-0.81	0.418	-.6178835	.2602468
Bonehealing						
2	.5973152	.4096912	1.46	0.150	-.2221892	1.41682
3	.2816708	.3360304	0.84	0.405	-.3904901	.9538317
4	.2717252	.367179	0.74	0.462	-.4627422	1.006193
5	-.16918	.8444604	-0.20	0.842	-1.858352	1.519992
1.BCereus	.9843949	.2231356	4.41	0.000	.5380572	1.430732
_cons	4.685803	.5934943	7.90	0.000	3.498637	5.872968

To compare to another category, in this example to category 4. Put b(4). rather than i. before Bonehealing. In this example, B.Cereus looks to be the only important influence on log time to closure.

```
. regress logTimeClose i.CHRONICINFECTION Age BMI i.Sex b(4).Bonehealing i.BCereus
```

Source	SS	df	MS	Number of obs	=	70
Model	18.5791296	9	2.06434773	F(9, 60)	=	3.04
Residual	40.7421598	60	.679035996	Prob > F	=	0.0046
				R-squared	=	0.3132
				Adj R-squared	=	0.2102
Total	59.3212893	69	.859728831	Root MSE	=	.82404

logTimeClose	Coef.	Std. Err.	t	P> t	[95% Conf. Interval]	
1.CHRONICINFECTION	.0823017	.2224124	0.37	0.713	-.3625892	.5271927
Age	-.0006995	.0055025	-0.13	0.899	-.0117062	.0103071
BMI	.0227162	.0172758	1.31	0.194	-.0118405	.057273
Sex						
M	-.1788183	.2194999	-0.81	0.418	-.6178835	.2602468
Bonehealing						
1	-.2717252	.367179	-0.74	0.462	-1.006193	.4627422
2	.32559	.5247338	0.62	0.537	-.7240339	1.375214
3	.0099457	.4678493	0.02	0.983	-.9258923	.9457836
5	-.4409052	.9053846	-0.49	0.628	-2.251944	1.370134
1.BCereus	.9843949	.2231356	4.41	0.000	.5380572	1.430732
_cons	4.957528	.729117	6.80	0.000	3.499077	6.415979

To include an interaction effect use ## to automatically include both main effects and the interaction effect. Using a single # means you must add in the main effects separately. For example:

```
. regress logTimeClose i.CHRONICINFECTION i.Sex i.CHRONICINFECTION#i.Sex i.BCereus BMI
```

```
. regress logTimeClose i.CHRONICINFECTION##i.Sex i.BCereus BMI
```

Source	SS	df	MS	Number of obs	=	70
Model	16.5328238	5	3.30656476	F(5, 64)	=	4.95
Residual	42.7884655	64	.668569774	Prob > F	=	0.0007
				R-squared	=	0.2787
				Adj R-squared	=	0.2223
Total	59.3212893	69	.859728831	Root MSE	=	.81766

logTimeClose	Coef.	Std. Err.	t	P> t	[95% Conf. Interval]	
1.CHRONICINFECTION	.1353685	.2605297	0.52	0.605	-.3850994	.6558365
Sex						
M	-.2078547	.239169	-0.87	0.388	-.6856497	.2699404
CHRONICINFECTION#Sex						
1#M	.0666915	.4699383	0.14	0.888	-.8721182	1.005501
1.BCereus	.9863938	.224936	4.39	0.000	.5370325	1.435755
BMI	.0233051	.0167869	1.39	0.170	-.0102307	.0568408
_cons	4.711488	.53271	8.84	0.000	3.647277	5.775698

Margins is a useful post-regression command. It provides the predicted outcome for the model at specified levels of covariates in the model. It is especially useful for understanding the nature of an interaction effect as it gives the predicted outcome at each combination of the interacting variables. For example:

```
. margins CHRONICINFECTION#Sex
```

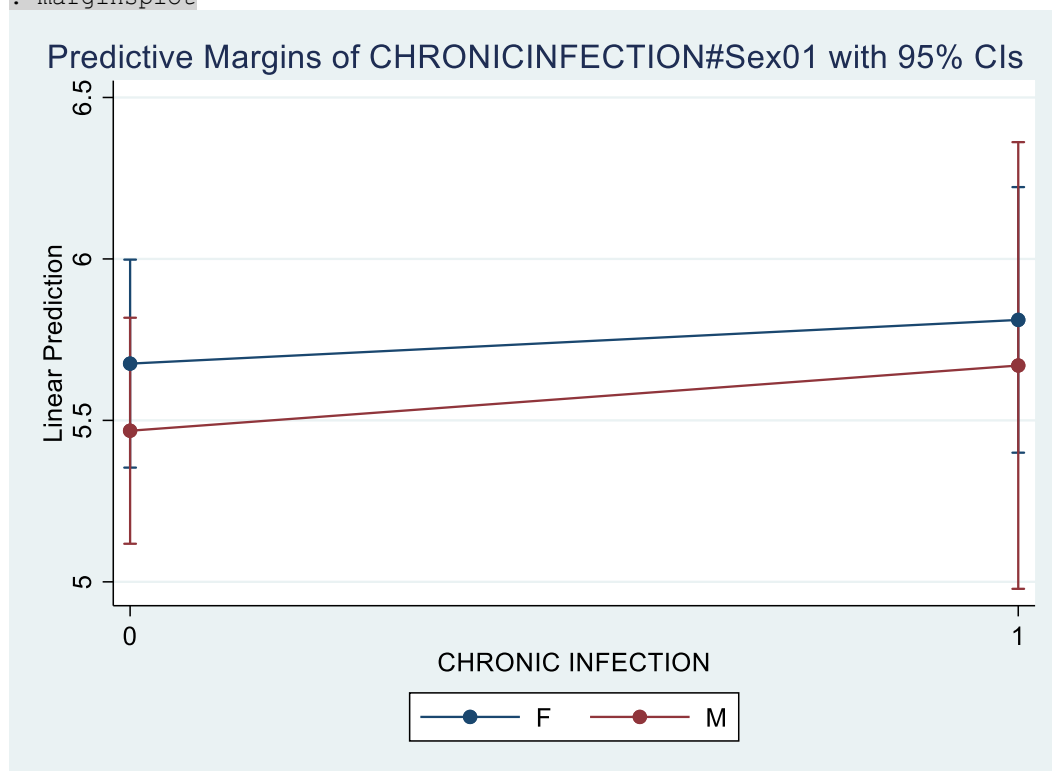
```
Predictive margins          Number of obs   =       70
Model VCE      : OLS
```

```
Expression      : Linear prediction, predict()
```

	Delta-method					[95% Conf. Interval]	
	Margin	Std. Err.	t	P> t			
CHRONICINFECTION#Sex							
0#F	5.675853	.1612051	35.21	0.000	5.353809	5.997897	
0#M	5.467998	.17518	31.21	0.000	5.118036	5.817961	
1#F	5.811222	.205839	28.23	0.000	5.400011	6.222432	
1#M	5.670058	.3462398	16.38	0.000	4.978365	6.361752	

Follow this up with marginsplot to graph the effect. Non-parallel lines suggest a significant interaction.

```
. marginsplot
```



In this example, with B.cereus:

```
. regress logTimeClose i.CHRONICINFECTION##i.BCereus BMI i.Sex
```

Source	SS	df	MS	Number of obs	=	70
Model	17.7822956	5	3.55645912	F(5, 64)	=	5.48
Residual	41.5389937	64	.649046777	Prob > F	=	0.0003
				R-squared	=	0.2998
				Adj R-squared	=	0.2451
Total	59.3212893	69	.859728831	Root MSE	=	.80563

	logTimeClose	Coef.	Std. Err.	t	P> t	[95% Conf. Interval]
1.CHRONICINFECTION						
1.BCereus		-.0506401	.258335	-0.20	0.845	-.5667235 .4654433
CHRONICINFECTION#BCereus						
1 1		.6453348	.4626284	1.39	0.168	-.2788717 1.569541
BMI		.0212396	.0165988	1.28	0.205	-.0119204 .0543996

```

Sex |
M | -.2509926 .2091696 -1.20 0.235 -.6688569 .1668717
cons | 4.845798 .531723 9.11 0.000 3.783559 5.908037
    
```

```
. margins CHRONICINFECTION#BCereus
```

```

Predictive margins          Number of obs   =       70
Model VCE      : OLS
    
```

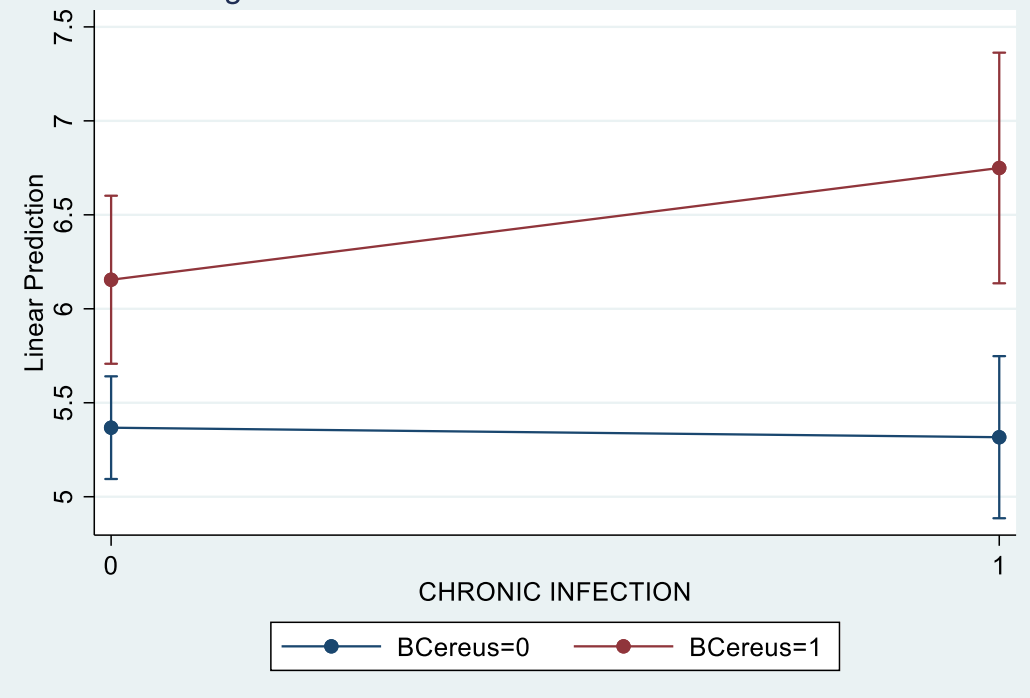
```
Expression   : Linear prediction, predict()
```

CHRONICINFECTION#BCereus	Delta-method				
	Margin	Std. Err.	t	P> t	[95% Conf. Interval]
0 0	5.367449	.1367317	39.26	0.000	5.094296 5.640602
0 1	6.154616	.2238037	27.50	0.000	5.707517 6.601715
1 0	5.316809	.2159119	24.62	0.000	4.885475 5.748143
1 1	6.749311	.3071336	21.98	0.000	6.135741 7.362881

```
. marginsplot
```

Variables that uniquely identify margins: CHRONICINFECTION BCereus

Predictive Margins of CHRONICINFECTION#BCereus with 95% CIs



Logistic regression can be performed with the logistic command, as below:

```
. logistic GoodResponse i.Sex BMI i.CHRONICINFECTION i.BCereus
```

```

Logistic regression          Number of obs   =       70
                             LR chi2(4)         =       44.67
                             Prob > chi2        =       0.0000
Log likelihood = -22.667119   Pseudo R2      =       0.4963
    
```

GoodResponse	Odds Ratio	Std. Err.	z	P> z	[95% Conf. Interval]
Sex					
M	1.887282	1.708759	0.70	0.483	.3200021 11.13066
BMI	.8892833	.058282	-1.79	0.073	.7820849 1.011175
1.CHRONICINFECTION	1.136505	.9736462	0.15	0.881	.2120044 6.092528
1.BCereus	.009316	.0093258	-4.67	0.000	.0013096 .0662717
_cons	220.1099	488.9732	2.43	0.015	2.829428 17123.02

Note: _cons estimates baseline odds.

Logistic regression is one of many generalized linear models and you can get the same result from



Statistics / Generalized linear models / Generalized linear model (GLM)

Then choosing the binomial family and the logit link.

```
. glm GoodResponse i.Sex BMI i.CHRONICINFECTION i.BCereus, family(binomial 1) link(logit)
```

```
Iteration 0: log likelihood = -23.418096
Iteration 1: log likelihood = -22.67379
Iteration 2: log likelihood = -22.66712
Iteration 3: log likelihood = -22.667119
```

```
Generalized linear models      No. of obs   =      70
Optimization      : ML          Residual df   =      65
                               Scale parameter =      1
Deviance           =  45.33423773 (1/df) Deviance = .6974498
Pearson           =  57.9795399   (1/df) Pearson = .8919929
```

```
Variance function: V(u) = u*(1-u)      [Bernoulli]
Link function      : g(u) = ln(u/(1-u)) [Logit]
```

```
Log likelihood = -22.66711886      AIC           = .7904891
                                           BIC           = -230.818
```

GoodResponse	Coef.	Std. Err.	z	P> z	[95% Conf. Interval]	
Sex						
M	.6351374	.9054071	0.70	0.483	-1.139428	2.409703
BMI	-.1173394	.0655382	-1.79	0.073	-.2457919	.0111131
1.CHRONICINFECTION	.1279574	.8567023	0.15	0.881	-1.551148	1.807063
1.BCereus	-4.676023	1.001055	-4.67	0.000	-6.638055	-2.713991
_cons	5.394126	2.221496	2.43	0.015	1.040074	9.748178

NOTE: These results are not exactly the same as when the logistic command was used. The results above report the coefficients (β s) of each predictor variable (the weightings of each predictor in the linear part of the model). The output following the logistic command gave the odds ratio. The odds ratio is the exponentiated form of the coefficient, e^{β} , which can be requested from the glm command by adding the eform option.

```
. glm GoodResponse i.Sex BMI i.CHRONICINFECTION i.BCereus, family(binomial 1) link(logit)
                                     eform
```

```
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```
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```
Variance function: V(u) = u*(1-u)      [Bernoulli]
Link function      : g(u) = ln(u/(1-u)) [Logit]
```

```
Log likelihood = -22.66711886      AIC           = .7904891
                                           BIC           = -230.818
```

GoodResponse	Odds Ratio	Std. Err.	z	P> z	[95% Conf. Interval]	
Sex						
M	1.887281	1.708758	0.70	0.483	.320002	11.13065
BMI	.8892833	.058282	-1.79	0.073	.7820849	1.011175
1.CHRONICINFECTION	1.136505	.9736461	0.15	0.881	.2120044	6.092527
1.BCereus	.009316	.0093258	-4.67	0.000	.0013096	.0662718
_cons	220.1097	488.9727	2.43	0.015	2.829426	17123

Note: _cons estimates baseline odds.

```
. set seed 22209  
. gen GoodResponse = runiform()  
. gen Bernouli = rbinomial(1, 0.5)  
. replace GoodResponse = 1 if GoodResponse>0.3  
(48 real changes made)  
. replace GoodResponse =0 if GoodResponse !=1  
(23 real changes made)
```